

## Research Article

# A New Methodology for Evaluation of Nematode Viability

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Nematodes infections are responsible for debilitating conditions and economic losses in domestic animals as well as livestock and are considered an important public health problem due to the high prevalence in humans. The nematode resistance for drugs has been reported for livestock, highlighting the importance for development of new anthelmintic compounds. The aim of the current study was to apply and compare fluorimetric techniques using Sytox and propidium iodide for evaluating the viability of *C. elegans* larvae after treatment with anthelmintic drugs. These fluorescent markers were efficient to stain larvae treated with ivermectin and albendazole sulfoxide. We observed that densitometric values were proportional to the concentration of dead larvae stained with both markers. Furthermore, data on motility test presented an inverse correlation with fluorimetric data when ivermectin was used. Our results showed that lower concentrations of drugs were effective to interfere in the processes of cellular transport while higher drugs concentrations were necessary in order to result in any damage to cell integrity. The methodology described in this work might be useful for studies that aim to evaluate the viability of nematodes, particularly for testing of new anthelmintic compounds using an easy, economic, reproducible, and no time-consuming technique.

## 1. Introduction

Intestinal nematode infections represent a debilitating threat for livestock and domestic animals worldwide as it may cause reduction in growth and weight gain and, depending on the severity of infection, it can lead to animal death. Moreover, production costs associated with treatment and control measures of parasitic infections can be very expensive [1]. In humans, the intestinal nematode infections (e.g., soil-transmitted helminthes) are highly prevalent and affect at least 1 billion people worldwide [2]. The control of nematodes is generally performed by the use of commercially available chemical compounds such as avermectins, benzimidazoles, and imidazothiazoles [3]. Despite the availability of several drugs for nematodes control, resistance to the main drugs has

been extensively reported for livestock, highlighting the need to found new anthelmintic compounds [4, 5].

*Caenorhabditis elegans*, a free-living bacteriovorous nematode, has been used as a good model for research new novel anthelmintics [6]. This nematode has been valuable in basic research on anthelmintic pharmacology of human and agricultural parasites [7] as well as at understanding the mechanisms of resistance to anthelmintics [8], mainly due to the phylogenetic relationship to other parasitic helminthes. Several characteristics make *C. elegans* a good model, such as cycle fast, easy laboratory maintenance, knowledge of its genome, and phylogenetic proximity to other nematodes [9–11]. Currently, a number of techniques are available for measuring *C. elegans* viability after treatment with known drugs or candidates, such as larval development assays

(ADLs, reproduction responses), or by the assessment of motility and colorimetric assays [12–15]. However, some of these techniques are time consuming and it can be subjective. According to Dickson and Gagnon [16], the discovery of new bioactive molecules is a long and expensive work, requiring investment of an average of 10 to 20 years and more than 200 million of dollars. Thus, the development of no subjective, economic, reproducible, and no time-consuming techniques would be useful.

Techniques that employ fluorescent markers such as propidium iodide and Sytox have been routinely used to measure cellular viability of mammalian cell [17]. Propidium iodide and Sytox also demonstrated to be efficient at staining nematode larvae [18, 19]. Sytox intercalates into nucleic acid and it is not able to passively cross the plasma membrane of viable cells [20], propidium iodide, which is also an intercalator to nucleic acid in cell death and, nevertheless, is also capable of going through intact cell membranes; however, it is expelled by viable cells [21]. Therefore, both markers stain nonviable cells. In this study, we applied and compared a feasible method, using the fluorescent markers propidium iodide and Sytox to evaluate the viability of *C. elegans* larvae after drug treatments.

## 2. Methodology

**2.1. *C. elegans* L<sub>3</sub> Production.** The strain of *C. elegans* was kindly provided by Professor Carlos Eduardo Winter (Universidade de São Paulo (USP)). L<sub>3</sub> larvae of *C. elegans* were grown on NGM plates 8P according to the methodology previously described [22, 23]. After seven days of culture in BOD incubator at 20°C, the plates were washed with M9 medium [22] and filtered through three sieves with pores of 40 μm, 30 μm, and 20 μm. L<sub>3</sub> larvae retained in the 20 μm strainer were collected by backwashing. The obtained larvae were washed by centrifugation at 700 g for 4 minutes, followed by two washes with M9 medium. Larvae average size was 527 μ (σ 3.4) long by 23.3 μ in diameter (σ 1,9) [24].

**2.2. Drug Tests with Fluorescent Markers.** In order to evaluate the proportion between the number of larvae (L<sub>3</sub>) and the fluorescence signal, initial concentration of 2000 L<sub>3</sub> larvae per well diluted in M9 medium was used for fluorimetric tests followed by 1:2 serial dilution of L<sub>3</sub> larvae per well. Briefly, 100 μL of larvae suspension was added to each well in a 96-well microplate, followed by immediate addition of 100 μL of the tested drugs solution (Albendazole sulfoxide, ivermectin, Sigma-Aldrich). Stock solutions (20 mM) for all drugs were prepared in M9 solution supplemented with 0.05% DMSO (Vetec, BR) and stored at -20°C protected from light. For albendazole sulfoxide, several concentrations (4000, 3000, 2000, 1000, 500, 100, 10, 1, 0.1, and 0.01 μM) were tested. Ivermectin was tested at these concentrations: 1000, 500, 100, 10, 1, 0.1, and 0.01 μM. The negative control was M9 solution with 0.05% DMSO, and methanol (CH<sub>3</sub>OH) was used as positive control at the following dilutions 50, 25, 5, 0.5, 0.05, and 0.005% [25, 26]. Quadruplicates were performed for each drug concentration and for controls. Incubation of

microplates with different drugs was performed in a BOD (incubator) at 20°C.

**2.3. Staining with Propidium Iodide and Sytox.** After 48 and 72 hours of incubation with the drugs, propidium iodide (Invitrogen, USA) and Sytox (Invitrogen, USA) were added to the microplates markers at a final concentration of 20 μM and 1 μM per well, respectively [18, 19]. Microplates were incubated for 15 minutes at room temperature in a horizontal shaker at 120 rpm followed by reading at LAS ImageQuant™ GE 4000 with excitation in white light and emission at 605 nm for propidium iodide and 575 nm for Sytox. Densitometric analyses of the images were performed using the software GE Image Quant TL 8.1. Images were taken at microscope (Leica DM500) 100x magnification: excitation at 510–560 nm and emission at 590 nm for propidium iodide, excitation at 450–490 nm and emission at 535 nm for Sytox using a capture system (Canon EOS 600D).

**2.4. Motility Test.** *C. elegans* L<sub>3</sub> were resuspended in M9, and then approximately 1000 larvae in 100 μL of suspension were added to each well in a 96-well microplate. Tested drugs were then added at the same concentrations described in fluorimetric methods. Microplates containing drugs and larvae were stored in BOD incubator at 20°C. After 48 and 72 hours, 10 μL of solution containing ca. 100 larvae was removed from each well for analysis and quantification of paralyzed larvae number using an optical microscope at 100x magnification. Larvae were considered paralyzed when presenting with straight body and absence of any motility [27].

**2.5. Statistical Analyses.** Data from densitometry and motility assays were tested by analysis of variance (ANOVA) and linear and nonlinear regression using the statistical program GraphPad Prism 5.0. The model was considered adequate to the data when  $r^2$  was above 0.8 for nonlinear regression and 0.95 for linear regression. Comparison of groups was performed using normality test of Kolmogorov-Smirnov, followed by two-way ANOVA; comparison of means was tested using Bonferroni correction test for multiple hypothesis. Correlation analysis was performed using Spearman rank correlation. Nonlinear regression analysis was used to calculate the IC<sub>50</sub> value.

## 3. Results

Both Sytox and propidium iodide were effective for staining larvae of *C. elegans* previously killed by treatment with 50% methanol (Figure 1), presenting a clear differentiation to viable larvae, which did not present any fluorescence (Supplementary Figure S1 in Supplementary Material available online at <http://dx.doi.org/10.1155/2015/879263>). When suspension of dead larvae was serially diluted and stained with both fluorescent markers (Figure 2(a)), the observed densitometric values were proportional to the concentration of dead larvae stained with Sytox (Figure 2(b)) and propidium iodide (Figure 2(c)). The comparison of the densitometric

TABLE 1: IC<sub>50</sub> drugs: ivermectin and albendazole sulfoxide, using different fluorimetric markers and motility test (means of times 48 and 72 hours). Different letters in the lines demonstrate that the means are different ( $P < 0.05$ ) according to Tukey test and  $t$ -test.

Drug	IC <sub>50</sub> P. iodide	IC <sub>50</sub> Sytox	IC <sub>50</sub> motility
Ivermectin	132.4 $\mu$ M $\pm$ 5.3 <sup>a</sup>	261.4 $\mu$ M $\pm$ 23.7 <sup>b</sup>	0.87 $\mu$ M $\pm$ 0.05 <sup>c</sup>
Albendazole sulfoxide	214.9 $\mu$ M $\pm$ 100.2 <sup>a</sup>	874.5 $\mu$ M $\pm$ 67.50 <sup>b</sup>	—

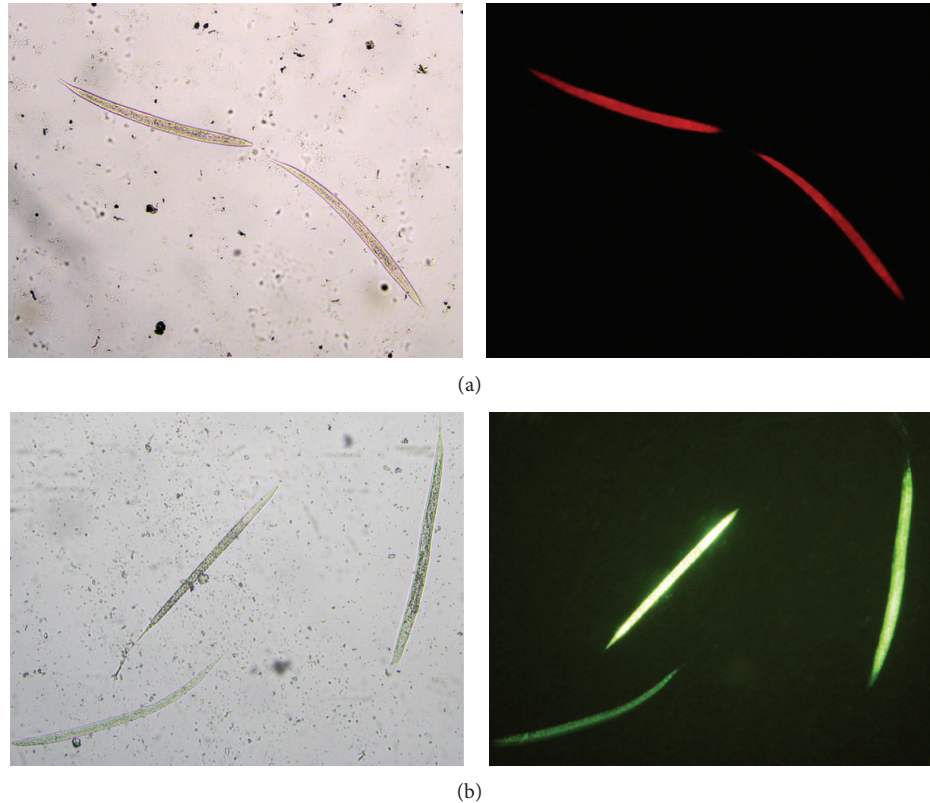


FIGURE 1: Dead *C. elegans* larvae (L<sub>3</sub>), bright field images on the left, and fluorescence images of same larvae on the right, propidium iodide (a) and Sytox (b).

values obtained by staining with Sytox and propidium iodide demonstrates that significant differences were observed only at high number of larvae (500 and 1000 larvae) ( $P < 0.05$ ) (Figure 2(d)).

Figure 3 shows the densitometric data of the larvae treated with albendazole sulfoxide and ivermectin and stained with propidium iodide. Our data suggest that the higher concentrations of drugs might affect the mechanisms used to expel the marker, once propidium iodide labels the nucleic acids when it remains within the cell. The densitometric data of larvae treated with the same drugs and stained with Sytox suggests that higher concentrations of drugs were able to induce damage in the cellular integrity of the larvae, because this marker is not able to overcome intact membranes (Figure 4). Concerning the ivermectin treatment, we observed that at the highest concentrations all larvae were practically inert (Figure 5). For instance, at concentration of 1000  $\mu$ M, 100% of paralysis was observed when compared to control group ( $P < 0.05$ ). A negative correlation (Spearman rank correlation,  $P < 0.05$ ) was observed when paralysis

and densitometry data were compared (Figure 6). The albendazole sulfoxide treatment induced a slightly reduction of the movement; however, body straight shapes and absence movement were not observed; thus, motility assay could not be properly performed (data not shown). The IC<sub>50</sub> dates are demonstrated at Table 1, where different IC<sub>50</sub> were observed. IC<sub>50</sub> data for albendazole sulfoxide motility test were not included once the larvae did not fit the criteria used for impairment of motility. Larvae were considered paralyzed when presenting straight body and absence of any motility; the treatment with albendazole sulfoxide induced reduction of the larval movement but body straight shapes and absence of movement were not observed.

#### 4. Discussion

In this study, we aimed to establish a methodology based on staining dead cells with the fluorescent markers Sytox and propidium iodide to evaluate the viability of *C. elegans* and

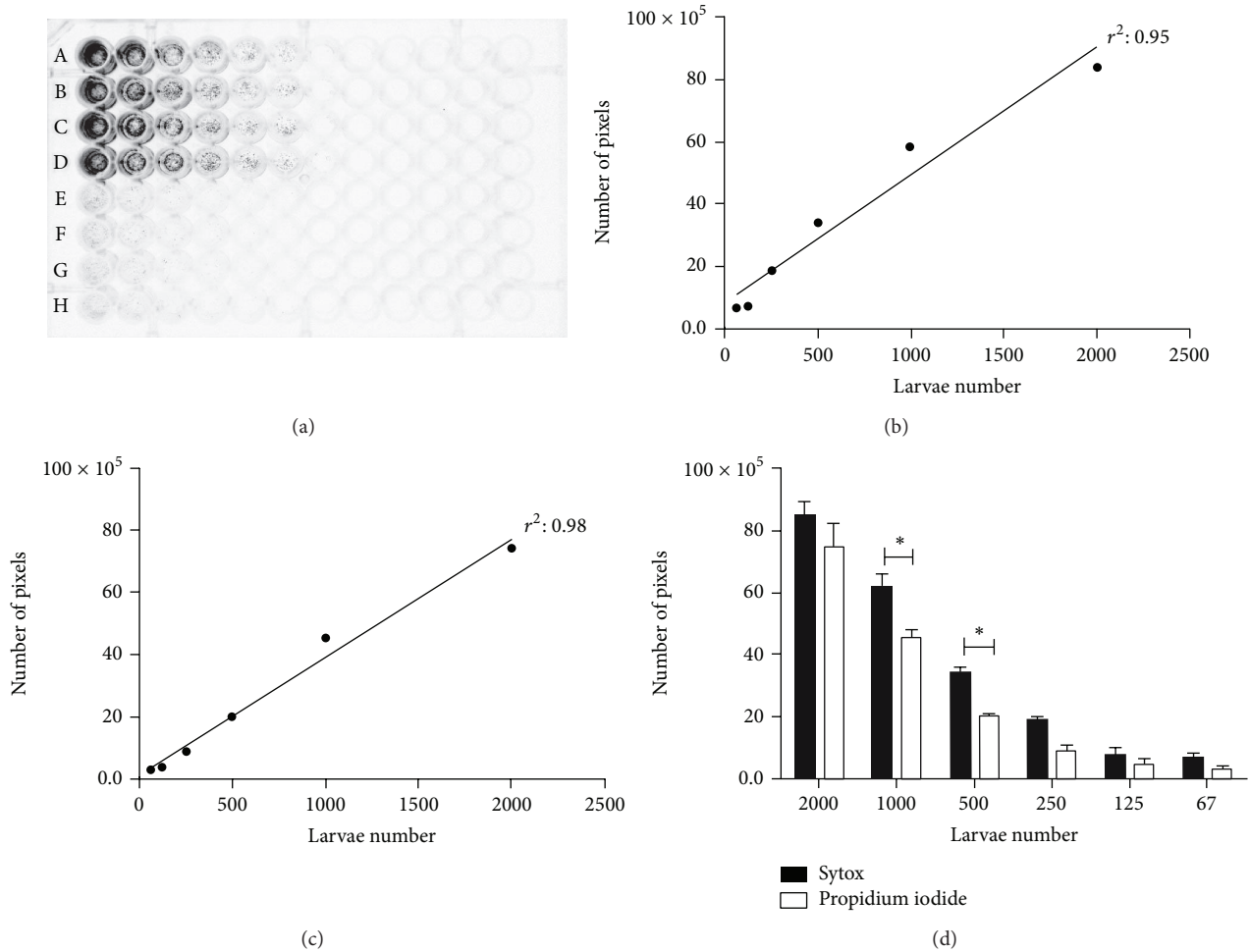


FIGURE 2: Analysis of viability using fluorimetric markers. (a) Microplate with serial dilution of *C. elegans* L<sub>3</sub> (first point 2,000 to 62). Rows A–D dead larvae; E–H represent viable (control) larvae. Correlation between the number of dead larvae with 50% methanol and the fluorescence intensity of the pixels with Sytox (b) and propidium iodide (c). Comparison of densitometry (number of pixels) between Sytox and propidium iodide markers (d). Significant differences ( $P < 0.05$ ) were only detected when 1000 and 500 larvae were tested.

its further use on testing of novel anthelmintic compounds for control of human and animal nematode infections.

During the standardization of the technique, the densitometric values observed for positive controls (larvae treated with 50% methanol, a concentration known to kill the larvae [25, 26]) were indeed proportional to the number of dead larvae, demonstrating the ideal number of larvae for the subsequent testing trials. Different number of larvae has been reported previously in the testing and standardization of new methodologies for several nematodes with a variety of IC<sub>50</sub> has been observed [14, 28, 29]. According to Smith et al. [30], such discrepancies may be attributed to differences in worm strain susceptibility against the toxic agent or variation in the methods employed in the analysis or effective drug concentration.

Once the standardization of number of larvae was performed, the staining methodologies were further tested with a representative compound of each main class of available drugs currently used. When ivermectin and albendazole

sulfoxide were used to induce worm death, we observed that both fluorimetric markers were efficient to stain dead larvae. Considering the mechanism of action of propidium iodide and Sytox [20, 21], our results allowed us to infer that treatment of larvae with ivermectin and albendazole sulfoxide may induce damage on the mechanism of exclusion of exogenous substances and/or affect the integrity of cell membranes. Indeed, the association of ivermectin association with glutamate-gated chloride channels (GluCl) results in the influx of chloride ion and consequent hyperpolarization of nematode muscle, culminating in the disarray on the locomotion mechanism and “pumping” of nutrients through the pharynx, promoting a flaccid paralysis and death of organism [31]. The albendazole sulfoxide is tubulin ligands with high and selective affinity for  $\beta$ -tubulin molecules, disturbing the microtubules polymerization and consequently preventing transport system mediated by microtubules [29].

Our results demonstrated that propidium iodide detected a smaller IC<sub>50</sub> for ivermectin and albendazole sulfoxide,

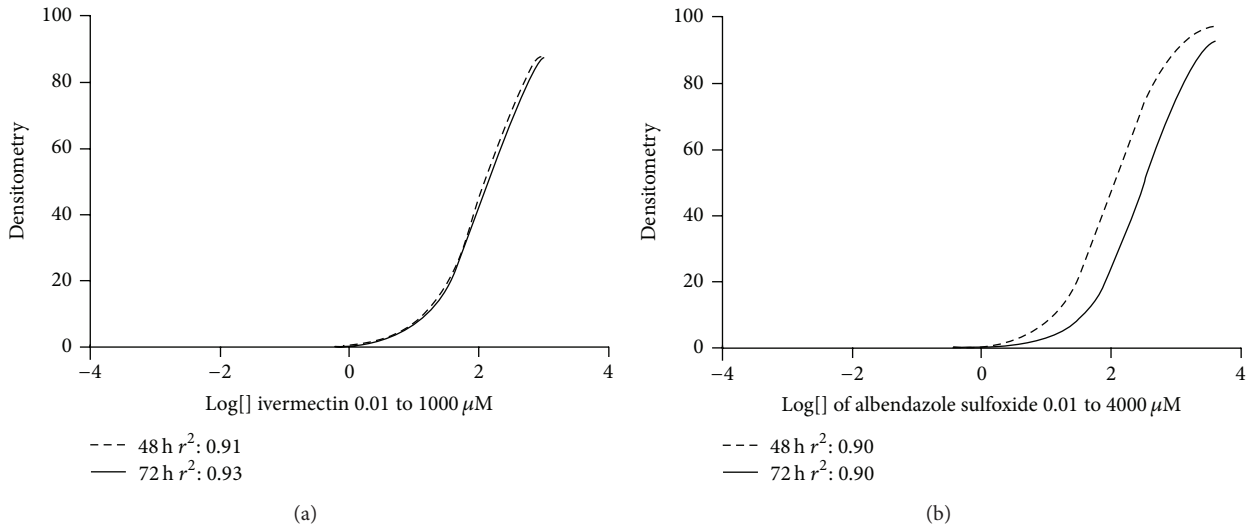


FIGURE 3: *C. elegans* larvae ( $L_3$ ) treated with ivermectin (a) at concentrations 0.01 to 1000  $\mu\text{M}$  (identical curves) and albendazole sulfoxide (b) at concentrations 0.01 to 4000  $\mu\text{M}$ , stained with propidium iodide. The larvae were incubated with the drugs for 48 and 72 hours in all treatment.

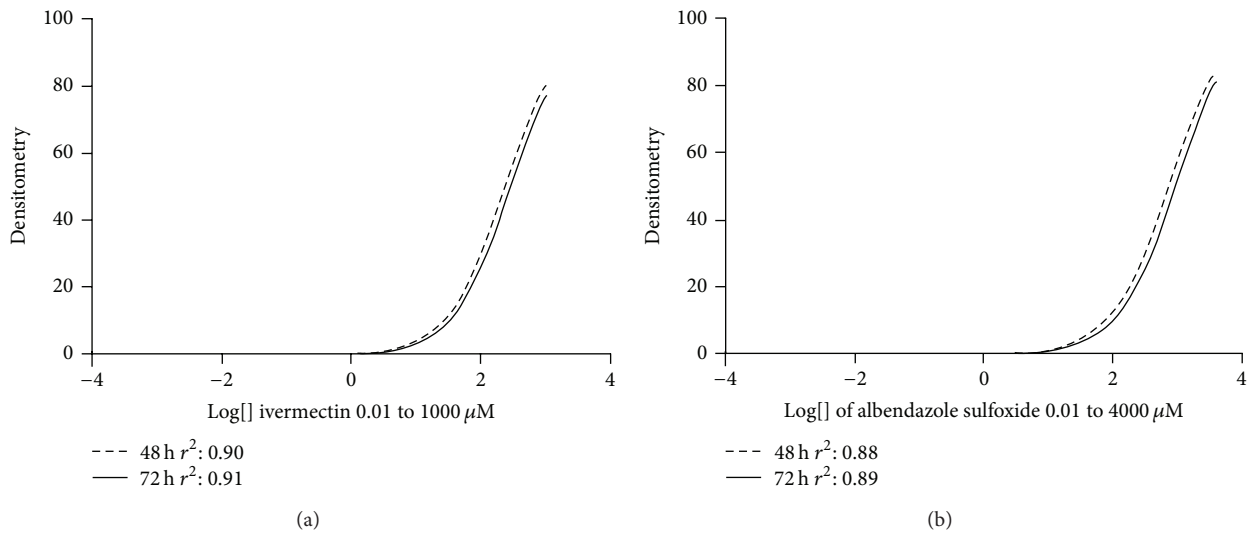


FIGURE 4: *C. elegans* larvae ( $L_3$ ) treated with ivermectin (a) at concentrations 0.01 to 1000  $\mu\text{M}$  and albendazole sulfoxide (b) at concentrations 0.01 to 4000  $\mu\text{M}$ , stained with Sytox. The larvae were incubated with the drugs for 48 and 72 hours in all treatment.

which is probably related to the cellular mechanism of influx or efflux responsible for excretion of several analytes including propidium iodide, suggesting that a relative lower drug concentration is sufficient to interfere with the mechanism of transport and the efflux of propidium iodide [29, 32, 33]. The ivermectin  $\text{IC}_{50}$  obtained with the motility assay was considerably lower than the  $\text{IC}_{50}$  obtained for fluorimetric assay, suggesting that a lower drug concentration is sufficient to impair the locomotion and disposition of worm body muscle as already described [27]. On the other hand, in order to disturb the mechanisms of exclusion of exogenous molecules or induce damage of cell membrane, higher concentrations of the drugs are required, and it is

known that used drugs in the present work can damage transport protein and cell membrane [32, 33]. Interestingly, the negative correlation between motility data and number of pixels in the fluorimetric assay indicates that ivermectin induces an efficient paralysis.

Taken together, our results show that fluorimetric microplate reading tests using propidium iodide and Sytox were efficient for larvae viability analysis after treatment with ivermectin and albendazole sulfoxide. This provides a simple and viable analysis technique to probe viability nematodes using *C. elegans* as a model, likely that is easy, less subjective, economic, reproducible, and no time consuming.

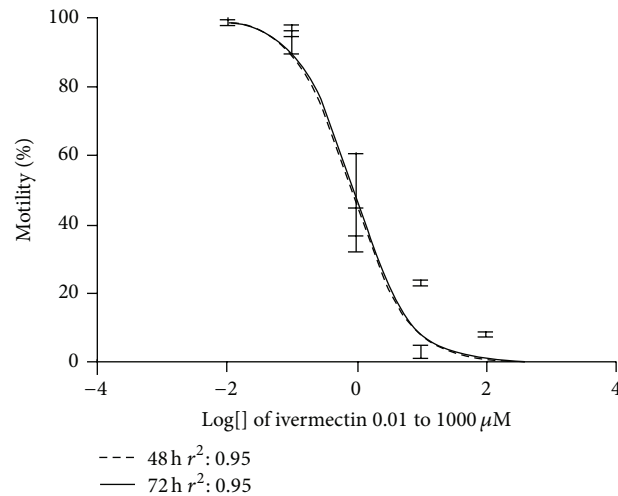


FIGURE 5: Motility analysis of *C. elegans* larvae ( $L_3$ ) treated with the ivermectin. The larvae were incubated with the drugs for 48 and 72 hours in all treatment.

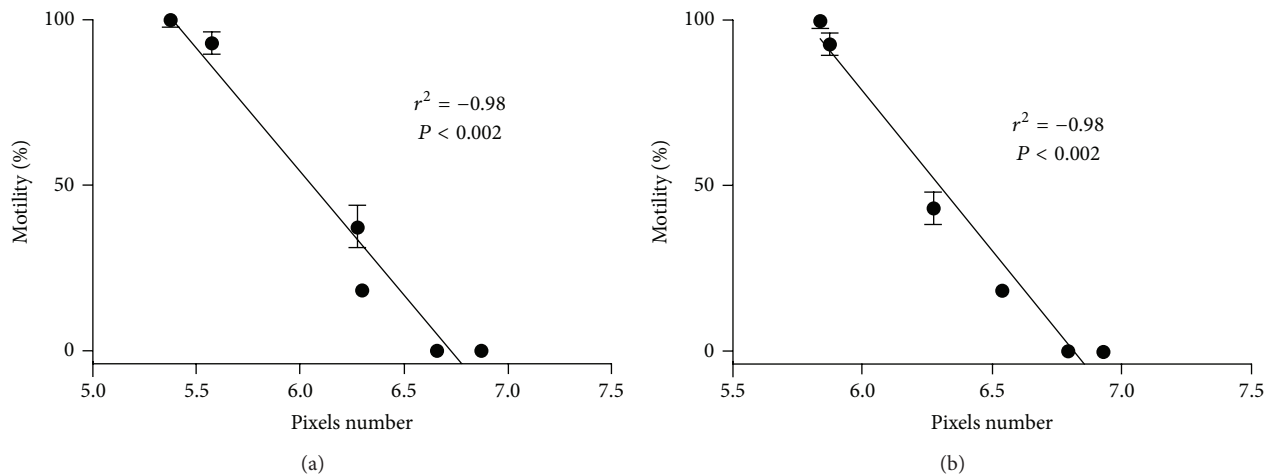


FIGURE 6: Correlation of paralysis and number of pixels from fluorimetric assays. (a) Propidium iodide and (b) Sytox. Statistical significance was determined by Spearman rank correlation.

## Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

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## References

- [1] F. Roeber, A. R. Jex, and R. B. Gasser, "Impact of gastrointestinal parasitic nematodes of sheep, and the role of advanced molecular tools for exploring epidemiology and drug resistance—an Australian perspective," *Parasites and Vectors*, vol. 6, no. 1, article 153, 2013.
- [2] R. L. Pullan, J. L. Smith, R. Jasrasaria, and S. J. Brooker, "Global numbers of infection and disease burden of soil transmitted helminth infections in 2010," *Parasites and Vectors*, vol. 7, no. 1, article 37, 2014.
- [3] A. J. Wolstenholme, I. Fairweather, R. Prichard, G. von Samson-Himmelstjerna, and N. C. Sangster, "Drug resistance in veterinary helminths," *Trends in Parasitology*, vol. 20, no. 10, pp. 469–476, 2004.
- [4] J. S. Gilleard, "Understanding anthelmintic resistance: the need for genomics and genetics," *International Journal for Parasitology*, vol. 36, no. 12, pp. 1227–1239, 2006.

- [5] C. J. Verissimo, S. C. M. Niciura, A. L. L. Alberti et al., "Multidrug and multispecies resistance in sheep flocks from São Paulo state, Brazil," *Veterinary Parasitology*, vol. 187, no. 1-2, pp. 209–216, 2012.
- [6] K. G. Simpkin and G. C. Coles, "The use of caenorhabditis-elegans for anthelmintic screening," *Journal of Chemical Technology and Biotechnology*, vol. 31, no. 1, pp. 66–69, 1981.
- [7] M. Dengg and J. C. A. van Meel, "Caenorhabditis elegans as model system for rapid toxicity assessment of pharmaceutical compounds," *Journal of Pharmacological and Toxicological Methods*, vol. 50, no. 3, pp. 209–214, 2004.
- [8] C. E. James, A. L. Hudson, and M. W. Davey, "Drug resistance mechanisms in helminths: is it survival of the fittest?" *Trends in Parasitology*, vol. 25, no. 7, pp. 328–335, 2009.
- [9] M. L. Blaxter, P. de Ley, J. R. Garey et al., "A molecular evolutionary framework for the phylum Nematoda," *Nature*, vol. 392, no. 6671, pp. 71–75, 1998.
- [10] T. G. Geary and D. P. Thompson, "Caenorhabditis elegans: how good a model for veterinary parasites?" *Veterinary Parasitology*, vol. 101, no. 3-4, pp. 371–386, 2001.
- [11] L. M. Katiki, J. F. S. Ferreira, A. M. Zajac et al., "Caenorhabditis elegans as a model to screen plant extracts and compounds as natural anthelmintics for veterinary use," *Veterinary Parasitology*, vol. 182, no. 2-4, pp. 264–268, 2011.
- [12] G. L. Anderson, R. D. Cole, and P. L. Williams, "Assessing behavioral toxicity with Caenorhabditis elegans," *Environmental Toxicology and Chemistry*, vol. 23, no. 5, pp. 1235–1240, 2004.
- [13] T. R. Bürglin, E. Lobos, and M. L. Blaxter, "Caenorhabditis elegans as a model for parasitic nematodes," *International Journal for Parasitology*, vol. 28, no. 3, pp. 395–411, 1998.
- [14] C. E. James and M. W. Davey, "A rapid colorimetric assay for the quantitation of the viability of free-living larvae of nematodes in vitro," *Parasitology Research*, vol. 101, no. 4, pp. 975–980, 2007.
- [15] J. B. Rand and C. D. Johnson, "Chapter 8 genetic pharmacology: interactions between drugs and gene products in Caenorhabditis elegans," *Methods in Cell Biology*, vol. 48, pp. 187–204, 1995.
- [16] M. Dickson and J. P. Gagnon, "The cost of new drug discovery and development," *Discovery Medicine*, vol. 4, pp. 172–179, 2004.
- [17] L. L. Chan, A. R. Wilkinson, B. D. Paradis, and N. Lai, "Rapid image-based cytometry for comparison of fluorescent viability staining methods," *Journal of Fluorescence*, vol. 22, no. 5, pp. 1301–1311, 2012.
- [18] P. R. Hunt, N. Olejnik, and R. L. Sprando, "Toxicity ranking of heavy metals with screening method using adult Caenorhabditis elegans and propidium iodide replicates toxicity ranking in rat," *Food and Chemical Toxicology*, vol. 50, no. 9, pp. 3280–3290, 2012.
- [19] M. S. Gill, A. Olsen, J. N. Sampayo, and G. J. Lithgow, "An automated high-throughput assay for survival of the nematode Caenorhabditis elegans," *Free Radical Biology & Medicine*, vol. 35, no. 6, pp. 558–565, 2003.
- [20] B. L. Roth, M. Poot, S. T. Yue, and P. J. Millard, "Bacterial viability and antibiotic susceptibility testing with SYTOX green nucleic acid stain," *Applied and Environmental Microbiology*, vol. 63, no. 6, pp. 2421–2431, 1997.
- [21] A. Nocker, C. Y. Cheung, and A. K. Camper, "Comparison of propidium monoazide with ethidium monoazide for differentiation of live vs. dead bacteria by selective removal of DNA from dead cells," *Journal of Microbiological Methods*, vol. 67, no. 2, pp. 310–320, 2006.
- [22] T. Stiernagle, *Maintenance of C. Elegans*, WormBook, 2006.
- [23] S. Brenner, "The genetics of Caenorhabditis elegans," *Genetics*, vol. 77, no. 1, pp. 71–94, 1974.
- [24] C. G. Knight, M. N. Patel, R. B. R. Azevedo, and A. M. Leroi, "A novel mode of ecdysozoan growth in Caenorhabditis elegans," *Evolution and Development*, vol. 4, no. 1, pp. 16–27, 2002.
- [25] D. Jones and E. P. M. Candido, "Feeding is inhibited by sublethal concentrations of toxicants and by heat stress in the nematode Caenorhabditis elegans: relationship to the cellular stress response," *Journal of Experimental Zoology*, vol. 284, no. 2, pp. 147–157, 1999.
- [26] G. Thompson and D. I. de Pomerai, "Toxicity of short-chain alcohols to the nematode Caenorhabditis elegans: a comparison of endpoints," *Journal of Biochemical and Molecular Toxicology*, vol. 19, no. 2, pp. 87–95, 2005.
- [27] I.-K. Park, J. Kim, S.-G. Lee, and S.-C. Shin, "Nematicidal activity of plant essential oils and components from Ajowan (Trachyspermum ammi), Allspice (Pimenta dioica) and Litsea (Litsea cubeba) essential oils against pine wood nematode (Bursaphelenchus xylophilus)," *Journal of Nematology*, vol. 39, no. 3, pp. 275–279, 2007.
- [28] L. Tritten, O. Braissant, and J. Keiser, "Comparison of novel and existing tools for studying drug sensitivity against the hookworm Ancylostoma ceylanicum in vitro," *Parasitology*, vol. 139, no. 3, pp. 348–357, 2012.
- [29] V. Sant'anna, R. C. Vommaro, and W. de Souza, "Caenorhabditis elegans as a model for the screening of anthelmintic compounds: ultrastructural study of the effects of albendazole," *Experimental Parasitology*, vol. 135, no. 1, pp. 1–8, 2013.
- [30] R. A. Smith, L. Pontiggia, C. Waterman, M. Lichtenwalner, and J. Wasserman, "Comparison of motility, recovery, and methylthiazolyl-tetrazolium reduction assays for use in screening plant products for anthelmintic activity," *Parasitology Research*, vol. 105, no. 5, pp. 1339–1343, 2009.
- [31] V. Portillo, S. Jagannathan, and A. J. Wolstenholme, "Distribution of glutamate-gated chloride channel subunits in the parasitic nematode Haemonchus contortus," *Journal of Comparative Neurology*, vol. 462, no. 2, pp. 213–222, 2003.
- [32] Y. N. Korystov, N. V. Ermakova, L. N. Kublik et al., "Avermectins inhibit multidrug resistance of tumor cells," *European Journal of Pharmacology*, vol. 493, no. 1–3, pp. 57–64, 2004.
- [33] M. H. Pourgholami, J. Akhter, L. Wang, Y. Lu, and D. L. Morris, "Antitumor activity of albendazole against the human colorectal cancer cell line HT-29: in vitro and in a xenograft model of peritoneal carcinomatosis," *Cancer Chemotherapy and Pharmacology*, vol. 55, no. 5, pp. 425–432, 2005.



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